

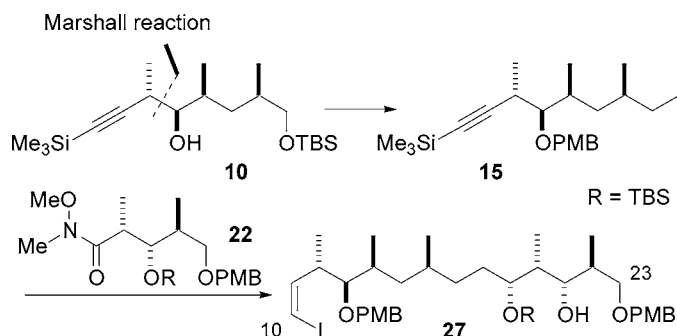
Chemoenzymatic Synthesis of the
C10–C23 Segment of Dictyostatin

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ABSTRACT



Employing enzymatic desymmetrization and resolution, respectively, the two aldehydes **8** and **20** were prepared. The precursor to aldehyde **8**, *meso*-2,4-dimethylglutaric anhydride **3**, could be obtained by base treatment of the diastereomeric mixture. Aldehyde **8** was extended to alkyne **10** by a Marshall reaction introducing four carbon atoms. Lithiation of the derived iodide **15** and trapping of the anion with amide **22** gave ketone **23**. This compound led to the C10–C23 fragment **27** of dictyostatin.

A range of cytotoxic natural products confer their biological activity through stabilization of the microtubule during cell division. The taxol and epothilone cases show that this mode of action can be of enormous clinical relevance.¹ Nevertheless, undesired toxicity or sensitivity toward the P-glycoprotein efflux pump can render compounds unsuitable for further clinical development. The polyketide dictyostatin (**1**) also turned out to be an inducer of tubulin polymerization (Figure 1). Most interestingly, it retains this activity in cells expressing the P-glycoprotein pump.² The 22-membered macrolide was originally isolated by Pettit and co-workers.³ Later, Wright et al. found the same compound in a different sponge.² Although the constitution was known, the stereochemistry was reported only recently by Wright and Paterson.⁴ A striking feature of dictyostatin is its structural

similarity to discodermolide. In fact, the 10 stereocenters in the overlapping regions all have the same configuration. This similarity was first recognized and exploited by Curran for the synthesis of discodermolide/dictyostatin hybrids.⁵ So far, total syntheses of dictyostatin have been achieved by the

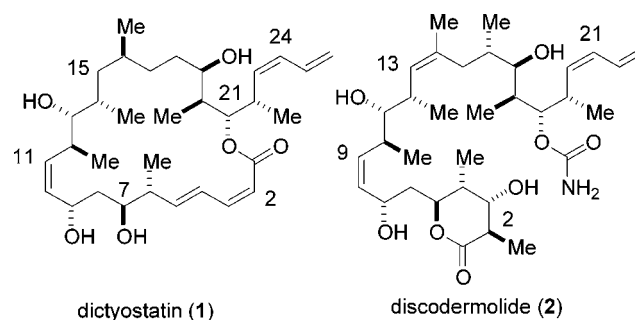


Figure 1. Structures of dictyostatin (**1**) and discodermolide (**2**).

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groups of Curran⁶ and Paterson.⁷ In addition, the synthesis of the C9–C19 subunit was described by Philipps.⁸ Biological studies are consistent with the hypothesis that the macrocyclic structure of dictyostatin resembles the bioactive conformation of the more flexible discodermolide.⁹ Most recently, an analogue, 16-normethyldictyostatin, turned out to have an activity profile toward several cell lines different from that of dictyostatin.¹⁰

The macrolactone of dictyostatin features a dienolate, a dienyl side chain, and several clusters of stereocenters that are commonly found in polyketides.^{11,12} Proven strategies to reach the stereotriads of discodermolide are based on aldol reactions with the Roche aldehyde^{13–16} or with methacrolein followed by hydroboration.¹⁷ To combine the various subunits, the Paterson group employed Wittig–Horner reactions. The Curran synthesis features an acetylide addition to a Weinreb amide and a Wittig–Horner coupling as key steps for connecting the subunits.

We devised a strategy to dictyostatin on the basis of an intramolecular Nozaki–Hiyama–Kishi reaction (Figure 2, structure **A**).^{18–20} Building block **B** containing a *Z*-vinyl iodide can be traced back to the 2,4-pentane diol **C** which is available from the corresponding diol by enzymatic sym-

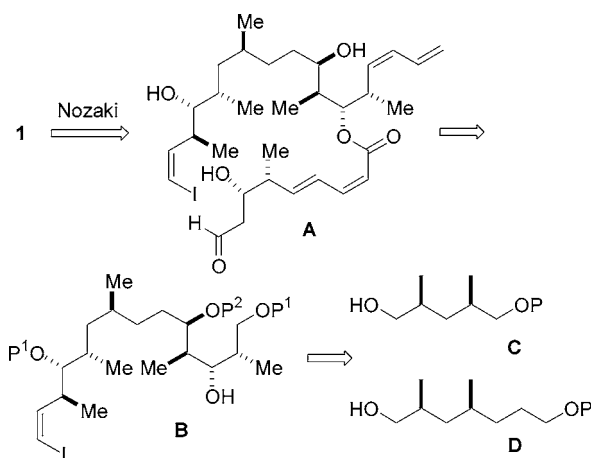


Figure 2. Retrosynthetic cuts for dictyostatin (**1**).

metry breaking.²¹ As an alternative, building block **D** could also be considered. Finally, we wanted to circumvent the use of the Roche ester because of its prohibitive costs. In this paper, we illustrate the realization of these goals.

The *meso*-diol **4** is available by reduction of the anhydride *meso*-**3** (Scheme 1). Unfortunately, the synthesis produces a mixture of diastereomeric anhydrides that has to be separated by repeated recrystallization leading to a low yield for *meso*-**3**.²² However, we found that the diastereomeric mixture of *meso*-**3**/*dl*-**3** could be converted more or less completely to *meso*-**3** by stirring the mixture with Hünig's base in ethyl acetate. One crystallization provided the *meso*-anhydride in excellent yield. The derived diol **4** was then converted to the acetate **5** using lipase Amano AK in the presence of vinyl acetate.²³ This reaction could be run on a multigram scale in high ee (98%). Silylation and basic cleavage of the acetate gave alcohol **7**.²⁴ This operation could be performed without chromatography. Oxidation²⁵ of alcohol **7** with bis(acetoxy)iodobenzene in the presence of catalytic amounts of tetramethyl-1-piperidinyloxy (TEMPO)

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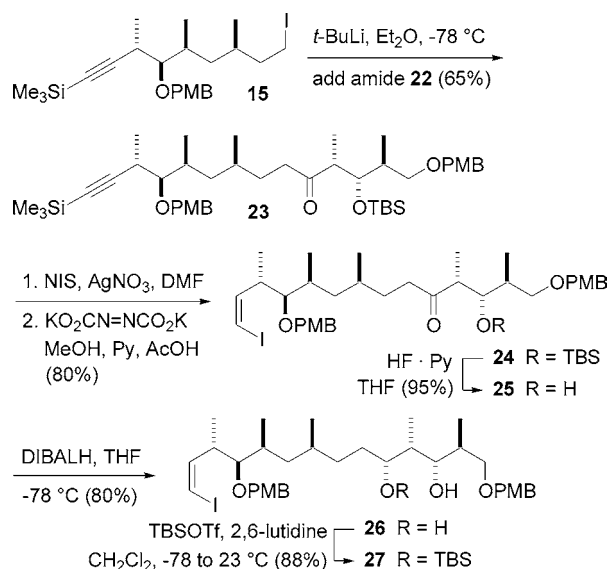
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Scheme 3. Combination of the Iodide **15** and the Weinreb Amide **22** and Transformation of the Resulting Ketone **23** to the C10–C23 Building Block **27**



Subsequently, the silyl ether was cleaved using the HF·pyridine complex.³⁴ Treatment of the hydroxy ketone **25** with DIBALH in THF induced a *syn*-selective reduction furnishing the 1,3-diol **26**.³⁵ Among the two hydroxyl groups of **26**, the left one (OH-19) is less hindered, and in fact, a

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selective silylation was easily possible,⁷ completing the synthesis of the C10–C23 segment **27**.

In summary, we could illustrate the use of *meso*-diol **4** for the synthesis of an important dictyostatin subunit. This diol is now easily available by equilibration of the diastereomeric dimethylglutaric anhydride. After enzymatic resolution of **4**, the derived aldehyde **8** was extended in one step to alkyne **10** via a Marshall reaction. Subsequently, homologation of **12** and coupling of the derived iodide **15** with Weinreb amide **22** led to ketone **23**. A *syn*-selective reduction on hydroxyketone **25** and a selective silylation completed the synthesis of the C10–C23 dictyostatin fragment **27**. Out of the eight stereocenters, five were ultimately derived by chemoenzymatic methods.

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Supporting Information Available: Experimental procedures and characterization for all new compounds reported and copies of NMR spectra for important intermediates. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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